# EXPERIMENTAL ARTICLES

# The Finding of the Yeast Species Saccharomyces bayanus in Far East Asia

G. I. Naumov<sup>\*,1</sup>, D. O. Gazdiev<sup>\*,\*\*</sup>, and E. S. Naumova<sup>\*</sup>

\*State Research Institute for Genetics and Selection of Industrial Microorganisms (GosNIIGenetika), Pervyi Dorozhnyi proezd 1, Moscow, 117545 Russia \*\*Botanical Garden, Amur Scientific Center, Far Eastern Division, Russian Academy of Sciences, Relochnyi per. 1, Blagoveshchensk, 675000 Russia Received April 15, 2002; in final form, June 26, 2003

**Abstract**—The genetic analyses of nine Far East Asian *Saccharomyces* isolates allowed us to identify three species (*S. cerevisiae, S. paradoxus*, and *S. bayanus*). The occurrence of the last species in Far East Asia was shown for the first time. A new methodology for the molecular genetic differentiation of *Saccharomyces* sensu stricto species is described. The ecogeographical distribution of *Saccharomyces* yeasts is discussed.

Key words: Saccharomyces, hybridization, PCR analysis, Far East.

Although the ecogeographical distribution of *Saccharomyces* sensu stricto yeasts has been studied for more than one hundred years, little is known about the ecogeography of its sibling *S. cerevisiae* species. The cosmopolitan species *S. cerevisiae* comprises mainly cultured strains, while its wild populations are few [1]. The related wild species *S. paradoxus* is widespread over the world and there are European, Far Eastern, Hawaiian, and North American divergent populations [2–5]. Three endemic wild *Saccharomyces* species, namely, *S. cariocanus* from Brazil and *S. kudriavzevii* and *S. mikatae* from Japan are also known [6].

Of particular interest is the cryophilic species *S. bayanus*, which is mainly associated with low-temperature fermentation processes in the production of wines and ciders in Europe [7–9]. The five known natural isolates of *S. bayanus* were recovered from the caddis fly *Mesophylax adopersus* in Spain (strain MCYC 623), the *Amanita citrina* mushroom in Slovenia (strain CCY 21-31-12), the fruit flies *Drosophila persimiles* in Yosemite National Park and *D. pseudoobscura* in the environs of Lake Berryessa, Davis, the United States (strains UCD 51-206 and UCD 61-137, respectively), and the hornbeam *Carpinus betulus* exudate in Hungary (strain NCAIM Y.00789) [1, 10].

The aim of this work was to identify the *Saccharomyces* strains isolated in the Amur oblast and the Primorsky Kray of Russian Far East and in North Korea.

### MATERIALS AND METHODS

Microbiological methods and strains. Saccharomyces yeasts were isolated from the exudates collected together with bark pieces from broad-leaved trees and from the fruits of relict plants (Table 1). The bark pieces were incubated at 28°C in a malt extract containing 4 ml/l of 40% lactic acid (chemically pure grade, Reakhim, Russia). The acid was added to the medium in order to inhibit bacterial growth. Samples taken from the malt extract in the phase of active fermentation (2 to 4 days of incubation) were plated onto malt extract agar, and the plates were incubated at 28°C for 2–3 days. The colonies grown on the malt extract agar plates were transferred onto YPD agar, a complete solid nutrient medium containing (g/l) yeast extract, 5; peptone, 10; glucose, 20; and agar, 20. After 1 day of incubation, the grown colonies were transferred onto a sporulation medium containing 30 g/l maltose and 20 g/l agar.

Genetic experiments were carried out using the complete YPD medium. Sporulation was induced by incubating the isolates in the acetate-containing medium described earlier [11]. Sugar fermentation was studied as described in the same paper [11]. Hybridization was carried out by the spore-to-spore method or by the mass mating of haploid cells on the complete YPD medium followed by the isolation of zygotes with a micromanipulator. The ascus walls were digested with the *Helix pomatia* snail gastric juice, and the ascospores were isolated with the micromanipulator.

The monosporous homothallic cultures of *S. baya*nus MCYC 623 (= CBS 7001), *S. cerevisiae* VKM Y-502 (= CBS 5287), and *S. paradoxus* CBS 5829 [1], marked by the mutations *ura3*, *ade1*, and *ade2*, respectively, were used as highly fertile standard test cultures. We also used the haploid genetic lines *S. cerevisiae* S288C (*gal2mal*) and X2180-1A (*agal2mal*). PCR analysis was carried out with the monosporous cultures

<sup>&</sup>lt;sup>1</sup> Corresponding author. E-mail: gnaumov@yahoo.com

Strain	Source, location, and year of isolation	Fermentation of:				Species
		Mel	Mal	Gal	Suc	affiliation
3.00	Grapes of the vine <i>Vitis amurensis</i> , Botanical Garden, Blagovesh- chensk, Amur oblast (2000)	_	-	+	+	S. cerevisiae
22.00	Fruits of the hawthorn <i>Grataegus dachurica</i> , Botanical Garden, Blagoveshchensk, Amur oblast (2000)	_	-	+	+	S. cerevisiae
136.01*	Exudate of the elm <i>Ulmus pumila</i> , Botanical Garden, Blagovesh- chensk, Amur oblast (2001)	+	+	+	+	S. bayanus
148.01*	Exudate of the elm <i>Ulmus pumila</i> , Botanical Garden, Blagovesh- chensk, Amur oblast (2001)	+	+	+	+	S. bayanus
159.01**	Exudate of the oak <i>Quercus mongolica</i> , Botanical Garden, Blagoveshchensk, Amur oblast (2001)	_	-	+	+	S. cerevisiae
163.01**	Exudate of the oak <i>Quercus mongolica</i> , Botanical Garden, Blagoveshchensk, Amur oblast (2001)	_	-	+	+	S. cerevisiae
61.02	Exudate of the aspen <i>Populus davidiana</i> , Marine Experimental Station, Khasan region, Primorsky Kray (2002)	_	-	+	+	S. paradoxus
CCY 21-4-89	Fermenting acorn meal, Pionsan, North Korea (1986)	-	+	+	+	S. cerevisiae
CCY 21-4-93	Sequoian leaves, Pionsan, North Korea (1986)	-	+	+	+	S. cerevisiae

Table 1. The origin and some characteristics of *Saccharomyces* sensu stricto strains

Note: Mel, Mal, Gal, and Suc stand for melibiose, maltose, galactose, and sucrose, respectively.

\* These strains were isolated from two trees 500 m apart.

\*\* These strains were isolated from two exudates of the same tree.

of S. cariocanus UFRJ 50816, S. kudriavzevii IFO 1802, and S. mikatae IFO 1815 as the control organisms. Along with the Saccharomyces strains that were isolated by us, we also identified two North Korean strains, CCY 21-4-89 and CCY 21-4-93 (Table 1) [12]. The culture collection name abbreviations used in this paper are as follows: MCYC, Departamento de Microbiologia, Escuela Tecnica Superior de Ingenieros Agronomos, Universidad Politecnica de Madrid, Spain; CBS, Centraalbureau voor Schimmelcultures, Utrecht, the Netherlands; VKM, All-Russia Collection of Microorganisms, Moscow, Russia; UFRJ, Instituto de Microbiologia, Universidade Federal do Rio de Janeiro, Brazil; IFO, Institute for Fermentation, Osaka, Japan; and CCY, Culture Collection of Yeasts, Institute of Chemistry, Slovak Academy of Sciences, Bratislava, Slovakia.

Molecular genetic methods. DNA was isolated as described earlier [13]. The 5.8S rDNA and internal transcribed ITS1 and ITS2 spacers (5.8S-ITS fragments) were amplified with the primers pITS1 (5'-TCCGTAGGTGAACCTGCGG-3') and pITS4 (5'-CCTCCGCTTATTGATATGC-3') [13] in 30 µl of a reaction mixture containing PCR buffer with 20 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 3 mM MgCl<sub>2</sub>, 0.25 mM of each dNTP, 0.3 µM of each primer, 1.25 U Taq polymerase (Sintol, Russia), and 20 ng of genomic DNA. PCR amplifications were run in a thermocycler manufactured by DNA Technology (Russia) with the initial DNA denaturation step at 94°C for 3 min, followed by 30 cycles of DNA denaturation at 94°C for 2 min, primer annealing at

MICROBIOLOGY Vol. 72 No. 6 2003

 $60^{\circ}$ C for 1 min, and DNA synthesis at 72°C for 2.5 min, with the final extension step at 72°C for 10 min. The amplification products were electrophoresed in 1% agarose gel at 60–65 V in 0.5× TBE buffer (45 mM Tris, 10 mM EDTA, and 45 mM boric acid) for 2 h. After electrophoresis, the gel was stained with ethidium bromide.

Restriction Fragment Length Polymorphism (RFLP) assay was carried out with *Hpa*II, *Hae*III, and *Taq*I restriction endonucleases purchased from Fermentas (Lithuania). The restriction digests were analyzed by electrophoresis in 1.6% agarose gel at 60–65 V in 0.5× TBE for 3 h. The gel was stained with ethidium bromide and photographed under UV light using a Vilber Lourmat transilluminator (France).

## **RESULTS AND DISCUSSION**

Seven Saccharomyces strains were isolated during the 2000–2002 expeditions in the south of Russian Far East. This territory, including the Amur oblast, is a unique ecosystem characterized by a combination of different zonal and exotic landscapes inhabited by endemic and relict plant and animal species. Here, several floral and faunal ecogeographical regions and provinces border. The yeast strains were isolated from the exudates of the oak *Quercus mongolica*, the elm *Ulmus pumila*, and the aspen *Populus davidiana* and from the fruits of relict arborifloral plants, the vine *Vitis amurensis* and the hawthorn *Grataegus dachurica* (Table 1). The genus affiliations of the isolates were



**Fig. 1.** The size of the amplified 5.8S–ITS rDNA fragments of the *Saccharomyces* sensu stricto strains. *S. cerevisiae*: (1) VKM Y-502, (2) 159.01, (3) 163.01, (4) 22.00, (5) 3.00; *S. paradoxus*: (6) CBS 5829, (7) 61.02; *S. bayanus*: (8) MCYC623, (9) 136.01, (10) 148.01; *S. kudriavzevii*: (11) IFO 1802; *S. mikatae*: (12) IFO 1815; *S. cariocanus*: (13) UFRJ 50816; M denotes the molecular weight marker 100-bp DNA Ladder (Fermentas, Lithuania).

determined by analyzing the morphology of their colonies, vegetative cells, and ascospores, as well as by the ability to ferment glucose [14]. The yeast species *S. bayanus* was differentiated based on the ability to ferment sugars, first of all, melibiose. Our studies confirmed the inability of *S. paradoxus* and the wild *Saccharomyces cerevisiae* strains to ferment maltose (Table 1).

The molecular genetic differentiation of the yeast isolates. Our earlier studies showed that six sibling species of the *Saccharomyces* sensu stricto complex differ in the sequence of the internal transcribed ITS1 and ITS2 rDNA spacers [6] and can be differentiated based on the analysis of the restriction fragment lengths of this region [13]. The species *S. cerevisiae, S. bayanus, S. paradoxus*, and *S. mikatae* can be differentiated with the aid of the *Hae*III and *Hpa*II restriction endonucleases, whereas the species *S. kudriavzevii* has a unique *TagI* restriction profile. Two species, *S. cariocanus* and *S. paradoxus*, do not differ in their RFLP



Fig. 2. The analysis of the amplified 5.8S–ITS rDNA fragments of the *Saccharomyces* sensu stricto strains with (a) *Hae*III and (b) *Hpa*II restriction endonucleases. *S. cerevisiae*: (1) VKM Y-502, (2) 159.01, (3) 163.01, (4) 22.00, (5) 3.00; *S. paradoxus*: (6) CBS 5829, (7) 61.02; *S. bayanus*: (8) MCYC623, (9) 136.01, (10) 148.01; *S. kudriavzevii*: (11) IFO 1802; *S. mikatae*: (12) IFO 1815; *S. cariocanus*: (13) UFRJ 50816; M denotes the molecular weight marker 100-bp DNA Lad.

profiles. The amplification of the 5.8S–ITS fragments of the seven Far Eastern isolates and six test cultures showed that their PCR products had the same size, 850 bp (Fig. 1), indicating that all of them belong to the *Saccharomyces* sensu stricto complex.

The analysis of PCR products with the HaeIII and *Hpa*II restriction endonucleases (Figs. 2a, 2b) showed that the yeasts studied can be divided into three groups according to their restriction fragment profiles. Group I included the test strain S. cerevisiae VKM Y-502 and four Far Eastern strains (3.00, 22.00, 159.01, and 163.01), which had four *Hae*III restriction fragments of about 320, 230, 170, and 130 kb in size (Fig. 2a, lanes 1-5) and two HpaII restriction fragments of about 730 and 120 kb in size (Fig. 2b, lanes 1–5. Group II included the test strain S. paradoxus CBS 5829, the Far Eastern strain 61.02, and S. cariocanus UFRJ 50816 (Figs. 2a, 2b, lanes 6, 7, and 13, respectively). The strains of group II were distinguished from the strains of group I by the absence of the HpaII restriction site, but had the same HaeIII restriction profiles. Group III included the test strains S. bayanus MCYC 623 and S. kudriavzevii IFO 1802 and two Far Eastern strains, 136.01 and 148.01 (Figs. 2a, 2b, lanes 8-11). The HaeIII restriction fragments of the members of this group had identical sizes (490, 230, and 130 kb). The HpaIII restriction profiles of this group were identical to those of the members of group I. The test strain S. mikatae IFO 1815 was similar to the strains of group III in the HaeIII restriction profile and to the strains of group II in the *Hpa*II restriction profile (Figs. 2a, 2b, lane 12). The TagI restriction profile of the test strain S. kudriavzevii IFO 1802 differed from the respective profiles of all other 12 strains (whose profiles were identical), suggesting that strains 136.01 and 148.01 belong to the species S. bayanus rather than to the species S. kudriavzevii. The results of RFLP assay showed that strain 61.02 belongs either to S. paradoxus or to S. cariocanus. In view of the fact that there are only two known strains of S. cariocanus, which are endemic to Brazil, while the species S. paradoxus is ubiquitous, the strain 61.02 was assigned to S. paradoxus.

Thus, the restriction enzyme analysis of the seven Far Eastern isolates showed that strains 3.00, 22.00, 159.01, and 163.01 belong to the species *S. cerevisiae*, strain 61.02 belongs to *S. paradoxus*, and strains 136.01 and 148.01 are referred to *S. bayanus*.

MICROBIOLOGY Vol. 72 No. 6 2003

Hybrid	Number of spore pairs crossed	Number of zygotes produced	Number of tetrads isolated	Viability of ascospores, %	Number of the 2ADE : 2ade tetrads analyzed
3.00 × 502	30	4	25	61	10
$22.00 \times 502$	30	2	25	80	12
$159.01 \times 502$	30	5	24	66	12
$163.01 \times 502$	30	9	24	65	8
$136.01 \times 502$	30	16	25	0	_
$148.01 \times 502$	30	8	25	0	-
$61.02 \times 502$	30	5	25	0	-
$3.00 \times 5829$	30	9	25	0	_
$22.00 \times 5829$	30	11	25	0	_
$159.01 \times 5829$	30	9	25	0	_
$163.01 \times 5829$	30	4	24	0	_
$61.02 \times 5829$	30	11	25	53	6
$136.01 \times 623$	30	14	25	99	18*
148.01 × 623	30	14	25	99	18*

**Table 2.** Analysis of the hybrids of *S. cerevisiae* (strains VKM Y-502, 3.00, 22.00, 159.01, and 163.01), *S. paradoxus* (strains CBS 5829 and 61.02), and *S. bayanus* (strains MCYC 623, 136.01, and 148.01)

Note: The asterisk marks the segregation 2URA : 2ura.

The genetic identification of the strains. Such studies require highly fertile monosporous yeast cultures. The analysis of the six to ten tetrads of the Far Eastern strains showed that they are homothallic and that their ascospores are highly viable (89–100%). The preliminary molecular genetic differentiation of the strains under study allowed us to minimize the number of hybridization experiments: each of these strains was crossed only with the suspected species and one of the control test cultures (either *S. cerevisiae* or *S. paradoxus*).

The ability of the Far Eastern strains to cross with the test cultures *S. cerevisiae*, *S. paradoxus*, and *S. bayanus* (Table 2) confirmed that they belong to the *Saccharomyces* sensu stricto complex. The high viability of hybrid ascospores and the regular meiotic segregation of the control markers *ade* and *ura* indicated a genetic homology of the parent strains and their affiliation to one species, whereas the sterility of hybrids suggested that their parents belong to different *Saccharomyces* species. The results of hybridization analysis confirmed the presumptive identification of the Far Eastern strains, namely, that strains 3.00, 22.00, 159.01, and 163.01 belong to the species *S. cerevisiae*, strain 61.02 belongs to *S. paradoxus*, and strains 136.01 and 148.01 belong to *S. bayanus* (Table 2).

The genetic study of the North Korean strains CCY 21-4-89 and CCY 21-4-93 was hampered by the low viability of their ascospores and the heterogeneous meiotic segregation of the strains with respect to the growth rate of colonies (including respiration-deficient microcolonies). The microscopic examination of monosporous cultures showed that they contain aggre-

gates of haploid cells. The mating types of monosporous clones were determined by the formation of zygotes in mixtures with the mating type testers S288C and X2180-1A. The strains CCY 21-4-89 and CCY 21-4-93 exhibited the meiotic segregations  $2\alpha$ :2a:3D and  $5\alpha$ :3**a**:1D, where  $\alpha$  and *a* denote haploid yeasts with the  $\alpha$  and **a** mating types and D denotes diploid sporulating yeasts. Fertile monosporous haploid clones were selected by analyzing their hybrids with the genetic lines S. cerevisiae S288C and X2180-1A (Table 3). Most of the hybrids were highly fertile but almost all of them, like the parent strains, exhibited heterogeneous segregation with respect to the growth rate of colonies. Based on the last characteristic, the two best monosporous cultures, 24-4-89:7A and 24-4-93:6B, were chosen for further analysis. The analysis of the ten tetrads of the hybrids of these cultures with strain S288C showed that these hybrids are characterized by the regular segregation 2 : 2 with respect to the control marker gal2. The hybrid of the strain 24-4-89:7A showed a monogenic segregation with respect to the marker mal. The other strain, 24-4-93:6B, likely had more polymeric MAL genes, as is evident from the analysis of the ten tetrads of its hybrid, eight of which exhibited the segregation 4Mal+:0Mal- and two of which exhibited the segregation 3Mal<sup>+</sup>:1Mal<sup>-</sup>. The results of this analysis showed that both North Korean strains belong to the species S. cerevisiae.

It should be noted that the ascomycetous yeasts of Russian Far East, including *Saccharomyces* sensu stricto, are far from being well studied [15], in contrast to the yeasts isolated in Japan [16–18]. Unfortunately,

Strain	Number of tet- rads analyzed	Viability of ascospores, %	Strain	Number of tet- rads analyzed	Viability of ascospores, %
21-4-89	9	44	24-4-93	8	44
21-4-89 : 7A × S288C	31	90	24-4-93 : 6B × S288C	28	96
21-4-89 : 7B × S288C	4	94	24-4-93 : 7B × S288C	6	88
21-4-89 : 8A × S288C	5	50	24-4-93 : 73 × S288C	7	96
21-4-89 : 9A × X2180-1A	5	100	21-4-93 : 8A × X2180-1A	7	96

**Table 3.** The viability of the ascospores of the North Korean strains CCY 21-4-89 and CCY 21-4-93 and their hybrids with the genetic lines *S. cerevisiae* S288C and X2180-1A

the ecogeographical studies of the Japanese isolates had been performed before the sibling species of *S. cerevisiae* were discovered. Our analysis of six wild Japanese *Saccharomyces* strains showed that four of them belong to the species *S. cerevisiae* and two strains belong to the species *S. paradoxus*. Four other strains were earlier described as new species *S. kudriavzevii* and *S. mikatae* [6].

Among the 17 wild *Saccharomyces* strains from Russian Far East, 5 were found to belong to the species *S. cerevisiae* and 12 were assigned to the species *S. paradoxus* [2, 20]. Four other wild *Saccharomyces* strains isolated from bordering areas (three Siberian strains and one Middle Asian strain) were also identified as *S. cerevisiae* strains.

Thus, in this work, we succeeded in the identification among the Far Eastern natural Saccharomyces isolates of three species, S. cerevisiae, S. paradoxus, and S. bayanus (the last species was detected in Far East for the first time). Noteworthy is the fact that both of the studied S. bayanus strains were isolated from the exudate of the elm Ulmus pumila (Table 1). Another known S. bayanus strain was isolated from the exudate of the hornbeam Carpinus betulus in Hungary [10]. At the same time, among the 50 Saccharomyces strains isolated from the exudates of the oaks *Quercus robur*, Q. mongolica, Q. rubra, and Q. alba in East Europe, the Caucasus, Far East, and North America, we were able to identify only the species S. paradoxus (46 isolates) and S. cerevisiae (4 isolates). These data suggest that the species S. bayanus is likely associated with the exudates of particular broad-leaved trees. Seasonal and climatic temperature factors likely play an important part in the ecogeographical distribution of cryophilic S. bayanus yeasts. The incubation of S. bayanus-containing enrichment cultures at low temperatures (below 20°C) may facilitate the isolation of this species. The active MEL-encoded  $\alpha$ ]-galactosidase of S. bayanus likely plays an adaptive role. It should be noted in this regard that the S. paradoxus strain isolated from the exudate of the elm Ulmus carpinifolia in the United States [3] is also able to ferment melibiose, which is a very rare ability for this species.

Based on the Vavilov's theory of geographical centers of biodiversity and taking into account that most of the Saccharomyces sensu stricto species (S. bayanus, S. cerevisiae, S. kudriavzevii, S. mikatae, and S. paradoxus) were found in Far Eastern Asia, we can suggest that it is expedient to search for new Saccharomyces species in this geographical region. The gene pool of the Far Eastern Saccharomyces isolates is of great importance for fundamental studies.

#### ACKNOWLEDGMENTS

This work was supported by grant no. 03-04-49245 from the Russian Foundation for Basic Research.

#### REFERENCES

- Naumov, G.I., Genetic Identification of Biological Species in the *Saccharomyces* sensu stricto Complex, *J. Ind. Microbiol.*, 1996, vol. 17, pp. 295–302.
- Naumov, G.I., Naumova, E.S., Azbukina, Z.M., Korhola, M., and Gaillardin, C., Genetic and Karyotypic Identification of *Saccharomyces* Yeasts from Far East Asia, *Cryptogamie Mycol.*, 1993, vol. 14, pp. 85–93.
- Naumov, G.I., Naumova, E.S., and Sniegowski, P.D., Differentiation of European and Far East Asian Populations of *Saccharomyces paradoxus* by Allozyme Analysis, *Int. J. Syst. Bacteriol.*, 1997, vol. 47, pp. 341–344.
- Naumov, G.I., Naumova, E.S., and Sniegowski, P.D., Saccharomyces paradoxus and Saccharomyces cerevisiae Are Associated with Exudates of North American Oaks, Can. J. Microbiol., 1998, vol. 44, pp. 1045–1050.
- Naumov, G.I., The Divergent Population of Saccharomyces paradoxus Yeasts in Hawaii: The Species in statu nascendi, Dokl. Akad. Nauk, 1999, vol. 364, no. 2, pp. 281–283.
- Naumov, G.I., James, S.A., Naumova, E.S., Louis, E.J., and Roberts, I.N., Three New Species in the Saccharomyces sensu stricto Complex: Saccharomyces cariocanus, Saccharomyces kudriavzevii, and Saccharomyces mikatae, Int. J. Syst. Evol. Microbiol., 2000, vol. 50, pp. 1931–1942.
- Naumov, G.I., Masneuf, I., Naumova, E.S., Aigle, M., and Dubourdieu, D., Association of *Saccharomyces bayanus* var. *uvarum* with Some French Wines: Genetic Analysis of Yeast Populations, *Res. Microbiol.*, 2000, vol. 151, no. 8, pp. 683–691.
- Naumov, G.I., Naumova, E.S., Aigle, M., Masneuf, I., and Belarbi, A., Genetic Reidentification of the Pectinolytic Yeast Strain SCPP As a *Saccharomyces bayanus*

var. uvarum, Appl. Microbiol. Biotechnol., 2001, vol. 55, pp. 108–111.

- Naumov, G.I., Nguyen, H.-V., Naumova, E.S., Michel, A., Aigle, M., and Gaillardin, C., Genetic Identification of *Saccharomyces bayanus* var. *uvarum*, a Cider-fermenting Yeast, *Int. J. Food Microbiol.*, 2001, vol. 65, pp. 163–171.
- 10. Naumov, G.I., *Saccharomyces bayanus* var. *uvarum* comb. nov., a New Variety Established by Genetic Analysis, *Mikrobiologiya*, 2000, vol. 69, no. 3, pp. 410–414.
- Naumov, G.I., Naumova, E.S., Korshunova, I.V., and Jakobsen, M., The Comparative Genetics of Yeasts: A New α-Galactosidase Gene *MEL15* in *Saccharomyces cerevisiae, Genetika*, 2002, vol. 38, no. 10, pp. 1330–1336.
- 12. Kockova-Kratochvilova, A., Slavikova, E., and Jan Do Sin, Yeasts and Yeast-Like Organisms from North Korea, *J. Gen. Appl. Microbiol.*, 1989, vol. 35, pp. 135–149.
- 13. Naumova, E.S., Korshunova, I.V., Jespersen, L., and Naumov, G.I., Molecular Genetic Identification of *Saccharomyces* sensu stricto Strains from African Sorghum Beer, *FEMS Yeast Res.*, 2003, vol. 3, pp. 177–184.
- Yarrow, D., Genus 22: Saccharomyces Meyen ex Reess, The Yeasts: A Taxonomic Study, Kreger-van Rij, N.Y.W., Ed., Amsterdam: Elsevier, 1984, pp. 379–395.

- 15. Bab'eva, I.P. and Reshetova, I.S., The Taxonomic Analysis of Yeasts from Russian Far East, *Mikol. Fitopatol.*, 1996, vol. 30, no. 4, pp. 10–18.
- Kodama, K., Ascosporogenous Yeasts Isolated from Tree Exudates in Japan, Ann. Microbiol. Enzimol., 1974, vol. 24, no. 2, pp. 215–231.
- Banno, I., Saccharomyces Yeasts Isolated in Japan: I. A Numerical Analysis of Saccharomyces cerevisiae and Its Allied Species, IFO Res. Commun., 1975, vol. 7, pp. 15–23.
- Banno, I. and Mikata, K., Ascomycetous Yeasts Isolated from Forest Materials in Japan, *IFO Res. Commun.*, 1981, vol. 10, pp. 10–19.
- Naumov, G.I. and Nikonenko, T.A., East Asia As a Putative Homeland of the Cultured Yeast Saccharomyces cerevisiae, Izv. Sib. Otd. Akad. Nauk SSSR, Ser. Biol. Nauki, 1988, no. 20, issue 3, pp. 97–101.
- Naumov, G.I., The Hybridological Study of Saccharomyces Yeasts Collected by V.I. Kudryavtsev during the 1934 and 1936 Expeditions, *Mikol. Fitopatol.*, 1988, vol. 22, no. 4, pp. 995–301.